CSIR-FRI/RE/BBA/2005/015



A Qualitative, Pathway-Initiated Risk Assessment

Bernard A. Boateng, Entomologist, University of Ghana, Legon, Ghana Haruna Braimah, Entomologist, Crops Research Institute, Kumasi, Ghana **Mary Glover-Amengor, Biochemist, Food Research Institute, Accra, Ghana** Angela Osei-Sarfo, Applied Biologist, Ghana Ministry of Food and Agriculture Ruth Woode, Entomologist, Ghana Ministry of Food and Agriculture Shawn Robertson, Entomologist, APHIS-IS Yu Takeuchi, Plant Physiologist, APHIS-PPQ CSIR-FRI/RE/BBA/2005/015

Importation of Okra, *Abelmoschus esculentus* from Ghana into the United States

A Qualitative, Pathway-Initiated Risk Assessment

Date June 2005

Agency contact: Shawn Robertson United States Department of Agriculture Animal and Plant Health Inspection Service Plant Protection and Quarantine Center for Plant Health Science and Technology Plant Epidemiology and Risk Analysis Laboratory 1017 Main Campus Drive, Suite 1550 Raleigh, NC 27606

Executive Summary

1

1

This risk assessment examined the risks associated with the importation of okra (*Abelmoschus esculentus*) from Ghana into the United States. Information on pests associated with okra in Ghana and neighboring countries revealed that six quarantine pests could potentially be introduced into the United States via this pathway. The quarantine pests likely to follow the pathway were all lepidopterous insects:

Cryptophlebia leucotreta (Meyrick) (Lepidoptera: Tortricidae) Earias biplaga Walker (Lepidoptera: Noctuidae) Earias insulana (Boisduval) (Lepidoptera: Noctuidae) Helicoverpa armigera (Hubner) (Lepidoptera: Noctuidae) Spodoptera littoralis (Boisduval) (Lepidoptera: Noctuidae) Leucinodes orbonalis Guenee (Lepidoptera: Pyralidae)

The quarantine pests were analyzed qualitatively based on international principles and internal guidelines as described in the PPQ Guidelines for Pathway-Initiated Pest Risk Assessments, Version 5.02 (USDA APHIS, 2000). This document examined pest biology in the context of Consequences of Introduction and Likelihood of Introduction. These elements were used to estimate the Pest Risk Potential. All of these pests pose phytosanitary risks to American agriculture. Port-of-entry inspections, as a sole mitigation measure, are considered insufficient to safeguard U.S. agriculture from all of these pests, and additional phytosanitary measures are necessary to reduce risks to acceptable levels.

Table of Contents

A. Introduction	3
B. Risk Assessment	3
1. Initiating Event: Proposed Action	3
2. Assessment of Weed Potential of Abelmoschus esculentus	3
Table 1. Assessment of the Weed Potential of Abelmoschus esculentus	4
3. Previous Risk Assessments, Current Status, and Pest Interceptions	4
4. Pest Categorization-Identification of Quarantine Pests and Quarantine Pests Likely	y to
Follow the Pathway	5
Table 2. Pests commonly associated with Abelmoschus esculentus in Ghana	5
5. Consequences of Introduction—Economic/Environmental Importance	14
Table 3. Risk Rating for Consequences of Introduction	20
6. Introduction Potential	21
2. Survive postharvest treatment:	22
3. Survive Shipment:	22
4. Not be detected at the port of entry:	22
5. Imported or move subsequently to an area with an environment suitable for sur	rvival:22
Table 4. Risk Rating for Likelihood of Introduction: (Risk Element #6)	23
C. Conclusion - Pest Risk Potential and Pests Requiring Phytosanitary Measures	23
Table 5. Summary of pest risk potential	23
D. Literature Cited	24

A. Introduction

This risk assessment was prepared for the Animal and Plant Health Inspection Service, (APHIS), U. S. Department of Agriculture (USDA) through a working group meeting of Ghanaian risk analysts, APHIS PPQ analysts and APHIS PPD analysts held in Accra, Ghana May 23-June 3, 2005. This working meeting was sponsored by the PRA advisor to the USAID West Africa Regional Program. The original risk assessment draft from which this one proceeded was completed by the Ministry of Food and Agriculture (MoFA) of Ghana as a result of training provided under an USDA/ICD/APHIS and Ghana PPQ Project [ATRIP Agricultural Grades and Standard Activity (PASA #641-P00-00-0042)].

This is a qualitative pest risk assessment that expresses risk in terms of high, medium, or low. Importing a new commodity gives exotic pests a potential pathway into the United States; this risk assessment is "pathway-initiated" in response to that threat.

International plant protection organizations, such as the North American Plant Protection Organization (NAPPO) and the International Plant Protection Convention (IPPC) of the United Nations Food and Agriculture Organization (FAO), provide guidance for conducting pest risk analyses. The methods used to initiate, conduct, and report this plant pest risk assessment are consistent with guidelines provided by NAPPO, IPPC, and FAO. Biological and phytosanitary terms (*e.g., introduction, quarantine pest*) conform with the NAPPO Compendium of Phytosanitary Terms (Hopper, 1995) and the Definitions and Abbreviations (Introduction Section) in International Standards for Phytosanitary Measures: Guidelines for Pest Risk Analysis (FAO, 1996).

FAO (1996) defines *pest risk assessment* as "determination of whether a pest is a quarantine pest and evaluation of its introduction potential." *Quarantine pest* is defined as "a pest of potential economic importance to the area endangered thereby and not yet present there, or present but not widely distributed and being officially controlled" (FAO, 1996; Hopper, 1995). Thus, pest risk assessments should consider both the consequences and likelihood of introduction of quarantine pests.

B. Risk Assessment

1. Initiating Event: Proposed Action

The USDA developed this risk assessment in response to a request by Ghana for a permit to import Okra (*Abelmoschus esculentus*) into the United States. The USDA has the authority to regulate imports of fruits and vegetables from foreign countries into the United States under Title 7, Part 319, Section 56 of the United States Code of Federal Regulations (7 CFR §319.56). The purpose of this risk assessment is to determine the likelihood that exotic plant pests would enter the United States with this commodity.

2. Assessment of Weed Potential of Abelmoschus esculentus.

This step examines the potential of the commodity to become a weed after it enters the United States (Table 1). If the assessment were to indicate significant weed potential, then a "pest-initiated" risk assessment would be conducted.

Table 1. Assessment of the Weed Potential of Abelmoschus esculentus

Commodity: Okra, Abelmoschus esculentus

Phase 1: Many varieties of *Abelmoschus esculentus* are widely cultivated in the United States. **Phase 2:** Is the species listed in:

- No Geographical Atlas of World Weeds (Holm et al., 1979)
- No World's Worst Weeds: Natural History and Distribution (Holm, 1997)
- No Report of the Technical Committee to Evaluate Noxious Weeds; Exotic Weeds for Federal Noxious Weed Act (Gunn and Ritchie, 1982)
- No Economically Important Foreign Weeds (Reed, 1977)
- No Weed Science Society of America list (WSSA, 1989)
- <u>No</u> Is there any literature reference indicating weed potential, *e.g.*, AGRICOLA, CAB, Biological Abstracts, AGRIS; search on "species name" combined with "weed." Such a search returns a prohibitive number of references, a sample of which do not indicate weed potential, but that there are weeds associated with the cultivation of *Abelmoschus esculentus*.

Phase 3: The literature indicates that *Abelmoschus esculentus* is not likely to become a weed in the United States because of imports from Ghana.

3. Previous Risk Assessments, Current Status, and Pest Interceptions

Decision History for Abelmoschus esculentus from Africa

1992. Okra from Nigeria. The request was denied because no approved treatment existed. The chief pests of concern were *Pectinophora gossypiella*, *Helicoverpa armigera*, and *Cryptophlebia leucotreta*.

1991. Okra from Senegal. The request was denied because of "Lack of pest data and quarantine treatment." The decision listed *Bactrocera ciliatus, Pectinophora gossypiella, Helicoverpa armigera,* and *Cryptophlebia leucotreta* as the chief pests of concern.

1989. Okra from Sierra Leone. The request was denied because of no treatments for *Earias insulana*. The decision also mentioned *Pectinophora gossypiella* as a pest of concern.

1989. Okra from Liberia. The request was denied because of "No acceptable treatment for a complex of exotic insect pests." The decision listed internal feeders such as *Earias insulana*, for which there was no treatment, and *Pectinophora gossypiella*.

Pest Interceptions

Between 1985 and 2004, U.S. agricultural inspectors intercepted numerous pests of okra, generally from passenger baggage (PIN, 2003). The following is a list of pests that were intercepted both on *Abelmoschus esculentus* from anywhere in the world and from Ghana on any commodity.

	Interception from
Interceptions on	West Africa on any
okra worldwide	commodity
1,602	8
101	40
95	11
30	79
16	38
	Interceptions on okra worldwide 1,602 101 95 30 16

		Interception from
	Interceptions on	West Africa on any
Pest	okra worldwide	commodity
Heliothis sp. (Lepidoptera: Noctuidae)	7	64
Cladosporium sp. (Fungi:)	7	25
Cryptophlebia sp. (Lepidoptera: Tortricidae)	6	1003
Aleurodicus dispersus Russell (Homoptera: Aleyrodidae)	5	8
Ceratitis capitata (Wiedemann) (Diptera: Tephritidae)	4	89
Helicoverpa armigera (Hubner) (Lepidoptera: Noctuidae)	4	4
Leucinodes orbonalis Guenee (Lepidoptera: Pyralidae)	4	3365
Diaphania sp. (Lepidoptera: Crambidae)	3	53
Spodoptera sp. (Lepidoptera: Noctuidae)	3	11

4. Pest Categorization-Identification of Quarantine Pests and Quarantine Pests Likely to Follow the Pathway

Common pests that are associated with *Abelmoschus esculentus* and occur in Ghana are listed in Table 2. This list includes information on the presence or absence of these pests in the United States, the affected plant part(s), the quarantine status of the pest with respect to the United States, pest-host association, and pertinent references for pest distribution and biology.

Pests identified only to genus or higher taxa were not considered for further analysis. Genera can contain many species; it is unrealistic to analyze an entire genus in which many species may not be pests. If pests identified only to higher taxa are intercepted in the future, the USDA may re-evaluate their risk. Intercepted pests are sometimes not identified to the species level because the current taxonomic knowledge is limited, the pest is too immature, or the specimen is in poor condition. By necessity, pest risk assessments focus on the organisms for which biological information is available. The lack of identification at the species level does not rule out the possibility that a high-risk quarantine pest was intercepted, or that the intercepted pest was not a quarantine pest. Conversely, detailed assessments for known pests that inhabit a variety of ecological niches, such as the surfaces or interiors of fruit, stems or roots, allow effective mitigation measures to eliminate the known organisms as well as similar, but incompletely identified organisms that inhabit the same niche.

and any abboe.		cumoscinits est	THE THEFT	II Omania	
West	US	Plant Part	Quaranti	Follow	References
African	Distribution	Affected ²	ne	Pathway	
Distribution ¹					а.
	5				
					<i>x</i>
SG	HIUS	F, L, S	No	Yes	CABI, 2004
				a a a a a a a a a a a a a a a a a a a	-
					,
	West African Distribution ¹	West US African Distribution Distribution ¹ US	West US Plant Part African Distribution Affected ² Distribution ¹ SG HI US F, L, S	West US Plant Part Quaranti African Distribution Affected ² ne Distribution ¹ SG HI US F, L, S No	West US Plant Part Quaranti Follow African Distribution Affected ² ne Pathway Distribution ¹ SG HI US F, L, S No Yes

Table 2. Pests commonly associated with Abelmoschus esculentus in Ghana

1 BF = Burkina Faso; BN = Benin; CI = Côte d'Ivoire; CM = Cameroun; CV = Cape Verde; FL = Florida; GH = Ghana; GM = Gambia; GU = Guinea; LB = Liberia; ML = Mali; MT = Mauritania; NG = Nigeria; NR = Niger; PR = Puerto Rico; SL = Sierra Leone; SN = Senegal; STP = Sao Tome & Principe; TG = Togo; US = United States; VI = Virgin Islands

2 F = Fruits; Fw = Flower; L = Leaves; = Roots; S = Stems; Sd = Seeds; W = whole plants (directly or indirectly as a result or crown or root destruction).

Pest	West African Distribution ¹	US Distribution	Plant Part Affected ²	Quaranti ne	Follow Pathway	References
Tetranychidae						8
Eutetranychus	CV NG SG		L	Yes	No	CABI, 2004
orientalis Klein						
Oligonychus	GH BE NG		L, S, W	Yes	No	Bolland et al., 1998
gossypii (Zacher)						
Tetranychus	CV TG	HIUS	F, L, S, W	No	Yes	CABI, 2004
cinnabarinus					A 20	1
(Boisduval)	014110					
Tetranychus sp.	GM'NG		F,L	Yes	Yes	PIN, 2003
Tetranychus	SL	US	L	No	No	CABI, 2004
urticae Koch						
Coleoptera						
Bruchidae						T
Spermophagus sp.	BF CV LB		Sd	Yes	No	PIN, 2003
Chrysomelidae					1	
Epilachna similis						
Thunberg						-
Podagrica	GB GH		L	Yes	No	CABI, 2004; Cobbinah
sjostedti Jacoby	NG SG					and Osei-Owusu, 1988;
						Vanlommel et al., 1996
Podagrica	GHNG		L	Yes	No	Obeng-Ofori and
uniformis (Jacoby)						Sackey, 2003;
						Vanlommel et al., 1996
Curculionidae						
Apion sp.	GM NG		F,Fw	Yes	No	PIN, 2003
	SN					
Dermestidae	1	1	-	_		
Trogoderma	ML NG SN		S	Yes	No	PIN, 2003
granarium Everts						
Elateridae						1
Conoderus sp.	LB		F	Yes	No ⁴	PIN, 2003
Meloidae			-			
Mylabris	GH		L	Yes	No	Obeng-Ofori and
temporalis						Sackey, 2003
Mylabris	GH		L	Yes	No	Obeng-Ofori and
trifasciata					2	Sackey, 2003
Scolytidae						
Hypothenemus sp.	CI GH LB		F	Yes	No	PIN, 2003
	NG					

3 Apion sp. was never intercepted on fruits from West Africa, intercepted only once on cutflowers from Ghana since 1985 PIN, 2003)

4 Not a known pest of okra in West Africa. There are no convincing published studies of this with supporting expert taxonomic identification (CABI, 2004).

Pest	West African Distribution ¹	US Distribution	Plant Part Affected ²	Quaranti ne	Follow Pathway	References
Tetranychidae						8
Eutetranychus	CV NG SG		L	Yes	No	CABI, 2004
orientalis Klein						
Oligonychus	GH BE NG		L, S, W	Yes	No	Bolland et al., 1998
gossypii (Zacher)						
Tetranychus	CV TG	HIUS	F, L, S, W	No	Yes	CABI, 2004
cinnabarinus					A 20	1
(Boisduval)	014110					
Tetranychus sp.	GM'NG		F,L	Yes	Yes	PIN, 2003
Tetranychus	SL	US	L	No	No	CABI, 2004
urticae Koch						
Coleoptera						
Bruchidae						T
Spermophagus sp.	BF CV LB		Sd	Yes	No	PIN, 2003
Chrysomelidae					1	
Epilachna similis						
Thunberg						-
Podagrica	GB GH		L	Yes	No	CABI, 2004; Cobbinah
sjostedti Jacoby	NG SG					and Osei-Owusu, 1988;
						Vanlommel et al., 1996
Podagrica	GHNG		L	Yes	No	Obeng-Ofori and
uniformis (Jacoby)						Sackey, 2003;
						Vanlommel et al., 1996
Curculionidae						
Apion sp.	GM NG		F,Fw	Yes	No	PIN, 2003
	SN					
Dermestidae	1	1	-	_		
Trogoderma	ML NG SN		S	Yes	No	PIN, 2003
granarium Everts						
Elateridae						1
Conoderus sp.	LB		F	Yes	No ⁴	PIN, 2003
Meloidae			-			
Mylabris	GH		L	Yes	No	Obeng-Ofori and
temporalis						Sackey, 2003
Mylabris	GH		L	Yes	No	Obeng-Ofori and
trifasciata					2	Sackey, 2003
Scolytidae						
Hypothenemus sp.	CI GH LB		F	Yes	No	PIN, 2003
	NG					

3 Apion sp. was never intercepted on fruits from West Africa, intercepted only once on cutflowers from Ghana since 1985 PIN, 2003)

4 Not a known pest of okra in West Africa. There are no convincing published studies of this with supporting expert taxonomic identification (CABI, 2004).

Pest	West African Distribution ¹	US Distribution	Plant Part Affected ²	Quaranti ne	Follow Pathway	References
Tenebrionidae						а. Т
Lagria cuprina			L	Yes	No	Obeng-Ofori and Sackey, 2003
Lagria villosa Fabricius	GH		L	Yes	No	Obeng-Ofori and Sackey, 2003
Coleoptera						
Scarabaeidae				¥2.		
Pachnoda interrupta (Olivier)	ML NG SG		F, R, Sd	Yes	No	CABI, 2004
Diptera						
Agromyzidae						_
<i>Liriomyza sativae</i> Blanchard	NG	HI US GU PR	L	No	No	CABI, 2004
<i>Liriomyza trifolii</i> <i>Burgess</i> in Comstock, 1880	BN CI GU NG SG	HI US VI GU PR	L	No	No	CABI, 2004
Cecidomyiidae		1				
Contarinia sp.	GHNG		F,Fw	Yes	No ⁵	PIN, 2003
Muscidae						
Atherigona orientalis Schiner	BF BN CV CI GH ML NR SL SG TG	HI US GU PR	F, Fw, L, Sd ,S, R, W	No	No	CABI, 2004
Tephritidae						
Ceratitis capitata (Wiedemann)	CV CI GM GH GU LB NG SN		F	Yes	No ⁶	PIN, 2003
Hemiptera			· · · · ·			
Aleyrodidae	1	-		1		E
<i>Bemisia tabaci</i> Gennadius	BF BN CV CI GM GH GU NG SL SG TG	HI US GU PR	L	No	No	CABI, 2004
Aphididae						
Aphis gossypii Glover, 1877	BF CV CI GM GU ML NR NG SL SG TG	HI US GU NMI PR	F,FW,S,W	No	No	CABI, 2004
<i>Myzus persicae</i> Sulzer (1776)	BN CI GH NG SL	HI US PR	Fw, L, S, W	No	No	CABI, 2004

⁵ Not a known pest of West Africa. Never intercepted on fruits from West Africa, intercepted only once on cut flowers from Ghana since 1985 PIN, 2003.

⁶ Not a known pest of okra. Only four insect interceptions but never from west africa PIN, 2003

Pest	West African Distribution ¹	US Distribution	Plant Part Affected ²	Quaranti ne	Follow Pathway	References
Cicadellidae						
Amrasca	GH	· · · ·	L, S	Yes	No	CABI, 2004; Obeng-
biguttula biguttula						Ofori and Sackey, 2003
Ishida =						
Empoasca					2	
davastans						
(Distant)						
Coccidae				1		
Parasaissetia nigra	GH		L, S	Yes	No	Ben-Dov, 1993; Ben-
(Nietner)						Dov <i>et al.</i> , 2004;
	OVCIOU	TH LIC M	T.C.	N	NL	CABI, 2004; Hill, 1994
Saissetia coffeae	NC SL TC	HIUS VI	L, S	NO	NO	CABI, 2004
(walker)	NG SL IG	GUPK				
Acmidialla	GU	LIC DD	EIS	No	Vac	CARL 2004
aurantii (Maskell)	00	USIK	I', L., 5	INO	105	CABI, 2004
Pseudaulacasnis	CVGH	US VI GU	LRSW	No	No	CABL 2004
nentagona	CV OII	PR	L, R, 5, W	110		C/IDI, 2004
(Targioni-		IR				
Tozzetti)						(A)
Pentatomidae			1		1	
Hotea subfasciata	GH		S	Yes	No	Leston, 1972
Nezara viridula	BF BN CV	HI US VI	F,Fw, Sd,	No	Yes	CABI, 2004
(Linnaeus)	CI GH GU	GU PR	R, S			
	ML NR					
	NG SL SG					
2	TG					
Pseudococcidae						
Ferrisia virgata	CI GH GU	HI US VI	F, L, S	No	No	CABI, 2004
(Cockerell)	NG SL SG	PR			1	
	TG					
Maconellicoccus	BF BN CI	HI US(FL)	F, Fw, L, S	Yes	No'	CABI, 2004
hirsutus (Green)	GM LB LB	VI GU PR				
	NR NG SG					
Phenacoccus sp.	NG		F	Yes	No	PIN, 2003

⁷ Never intercepted on fruits from West Africa, intercepted only thrice on fruits from the Caribbean PIN, 2003.

⁸ Easy to detect and cull infested fruit CABI, 2004.

Pest	West African	US Distribution	Plant Part Affected ²	Quaranti ne	Follow Pathway	References
Pvrrhocoridae	Distribution					
Dysdercus	GH, NG		Fw, Sd	Yes	No	Obeng-Ofori and
superstitiosus				1		Sackey, 2003
(Fabricius)						
Lepidoptera		100			90 - K	5
Gelechiidae			1			
Pectinophora	BF BN CI	HI US VI	F, Fw	No	Yes	CABI, 2004
gossypiella	GH ML	NMI PR				
(Saunders)	NK NG SL					
Noctuidae	5010					
Agrotis insilon	BF BN CI	HLUS	FISW	No	No	CABL 2004
Hufnagel	LBLBML	in es	1, 1, 5, 1		110	C/1D1, 2004
Thumager	SG TG					
Agrotis segetum	BN CV CI		L, R, S	Yes	No	CABI, 2004
(Denis &	ML SG TG					
Schiffermuller)						
Anomis flava	GH	US	L	No	No	CABI, 2004; Hill,
Fabricius	2					1994; Pogue, 2004;
1						Zhang, 1994
Earias biplaga	BF BN CI		F, Sd, W	Yes	Yes	CABI, 2004
Walker	GHGU	an ing Property State				
	ML NK					
Farias insulana	REPNCI		E Envil S	Vac	Vas	CAPI 2004
(Boisduval)	GHGU		1,1 W,L, 5	105	105	CADI, 2004
(Doisduvai)	MINR					Hard States
- Top and a set of the	NG SL SG		and the second second			
	TG					
Earias sp.	GH		F	Yes	Yes	PIN, 2003
Helicoverpa	BF BN CV		F, Fw, L	Yes	Yes	CABI, 2004
armigera	CI GM GH	and the latest state	A STREET			
(Hübner)	GUML					
	NR NG SL	designed and the second second		and the second		
	SG TG					
Helicoverpa sp.	GH		F	Yes	Yes	PIN, 2003
Spodoptera exigua	BF BN CI	HIUS	F, Fw, L	No	Yes	CABI, 2004
(Hubher)	MI ND SC					
· .	TG			1.0		
Spodontera	BEBNCV		FI	Vec	No	CABL 2004
littoralis	CLGMGH		1,1	105	140	CAD1, 2004
(Boisduyal)	GUML					
() = = = = = (, , , ,)	NR NG SL					
	SG TG					
Trichoplusia ni	CV GM	HI US VI	L, W	No	No	CABI, 2004
(Hübner)	NG SG	PR				

Pest	West African Distribution ¹	US Distribution	Plant Part Affected ²	Quaranti ne	Follow Pathway	References
Pyralidae						
Haritalodes	BF BN CI		L	Yes	No	CABI, 2004
derogata	GH ML					
(Fabricius)	NR NG SL				1	
	SG TG				<i>r</i>	
Leucinodes	GH SL NG		F	Yes	Yes	CABI, 2004
orbonalis Guenee						
Pyralidae sp.	GH		F	Yes	Yes	PIN, 2003
Sylepta derogata	GH		L	Yes	No	Cobbinah and Osei-
						Owusu, 1988; Obeng-
						Ofori and Sackey, 2003
Tortricidae						
Cryptophlebia	BF BN CV		F, L, Sd	Yes	Yes	CABI, 2004
leucotreta	CI GM GH					
(Meyrick)	MLNR					
	NG SL SG					
	TG					
Cryptophlebia sp.	GH NG TG		F	Yes	Yes	PIN, 2003
Orthoptera					1	
Acrididae						-
Diabolocatantops	BF BN CV		F.Fw.L. R.	Yes	No	CABI, 2004
axillaris	GH GU					
(Thunberg)	ML NR					
(NG SG					
Zonocerus	BF BN		F. Fw. L. S.	Yes	No	CABL 2004: Obeng-
variegatus (L.)	Chad CV		Sd. W			Ofori and Sackey 2003
()	GHGULB		00, 11		· · · · ·	ororraina Saokey, 2005
	ML NR					10-1
	NG SG SL				1 No. 1	
	TG				N	
Thysanoptera					1	
Thripidae					1	
Scirtothrips	GH		L, S	Yes	No	PIN, 2003
aurantii Faure					1	
Thrips	NG SL	HI US GU	F.Fw, L	No	Yes	CABI, 2004
hawaiiensis	-					
(Morgan)					à.	
Nematode			-1			-L
Hoplolaimidae						
Helicotylenchus	BF CI LB	HI US PR	L. R. W	No	No	CABI. 2004
dihystera (Cobb)	LB NG SG					
Sher						
Scutellonema	CI GM GH	US PR	R	No	No	CABI, 2004
bradys (Steiner &	GU NG SG					
Lehew) Andrassy	TG					

Pest	West	US	Plant Part	Quaranti	Follow	References
	Atrican	Distribution	Allected	ne	1 alliway	
Scutellonoma	Distribution BE BN CI		IPW	Ves	No	CABL 2004
clathricandatum	MINR		L, IX. W	105	INU	CADI, 2004
Whitehead	NG SI			1. A.		
Rotylenchulus	GH	US	R	No	No	Potter and Olthof
reniformis Linford	GII	00	in the second se			1990CABL 2004
& Oliveira						155001101, 2001
Longidoridae						
Xinhinema	CIGULB		L. R. W	Yes	No	CABI. 2004
<i>ifacolum</i> Luc	LBSL		2, 19, 11	1.00		
Meloidogynidae						
Meloidogyne	CI GM GH	HI US PR	L. R. W	No	No	CABI, 2004
arenaria (Neal)	LB LB NG		_,_,			
Chitwood	SG					
Meloidogyne	BF CI GM	HI US PR	L, R, W	No	No	CABI, 2004
incognita (Kofoid	GH GU LB					
& White)	LB NR NG					Р. е.,
Chitwood	SG					
Meloidogyne	CI GM GH	HI US PR	L, R, W	No	No	CABI, 2004
javanica (Treub)	LB LB NG					
Chitwood	SG					
Pratylenchidae					×	
Hirschmanniella	CI GM GH	US	L, R, W	No	No	CABI, 2004
oryzae (van Breda	GU NR					
de Haan)	NG SL SG				5. E	
Luc&Goodey						
Pratylenchus	BN CI GM	HI US PR	L, R, Sd, W	No	No	CABI, 2004
brachyurus	GU NG SG					
(Godfrey) Filipjev	TG		R.			
& Schuurmans					ing to the	
Stekhoven					1.1	
Pratylenchus loosi	SG		L, R. S. W	Yes	No	CABI, 2004
Loof. (Tylenchida:						
Pratylenchidae)					1	
Pratylenchus	NG	US	L,R,W	No	No	CABI, 2004
penetrans (Cobb)						
Fungi		1		1	1	
Alternaria	CI SG	HI US PR	F, Fw, L,	No	Yes	CABI, 2004
brassicae (Berk.)			Sd.S, W			
Sacc.						
Armillaria mellea	GH	US	R	No	No	Oduro, 1998
(Vahl) P. Kumm.						
Aspergillus niger	BF CI GU	US PR	F, Fw, L,	No	Yes	CABI, 2004
Tiegh.	NR NG		Sd. S, R, W			

Pest	West African Distribution ¹	US Distribution	Plant Part Affected ²	Quaranti ne	Follow Pathway	References
Botryodiplodia theobromae (Pat.) Griffiths & Maubl. [anamorph]	West Africa	US	F	No	Yes	CABI, 2004; Oduro, 1998
Cercospora hibiscina	GH		L	Yes	No	Oduro, 1998
<i>Cercospora</i> <i>malayensis</i> F.Stev. & Solh.	GH		L	Yes	No	CABI, 2004; Oduro, 1998
Cercospora sp.	GH	US	L	No	No	Oduro, 1998
Choanephora cucurbitarum (Berk. & Ravenel)	BN NG SG	US PR	F,Fw,Sd,L, S,W	No	Yes	CABI, 2004; Oduro, 1998
Cladosporium sp.	Chad GH GU LB NG SN		F	Yes	No ⁹	PIN, 2003
Cochliobolus lunatus R.R. Nelson & Haasis	BF BN GH NR NG	HI US PR	Fw, L, Sd	No	Yes	CABI, 2004
Colletotrichum sp.	GH GU LB NG		F	Yes	Yes	PIN, 2003
<i>Colletotrichum</i> <i>dematium</i> (Pers.) Grove	BF NR NG	US PR	F	No	Yes	CABI, 2004
Colletotrichum lindemuthianum (Sacc. & Magnus) Briosi & Cavara	GH	US	F S Sd	No	Yes	Gonzalez-Chavira et al., 2004; Oduro, 1998
Corynespora cassiicola	GH	US	L, Sd	No	Yes	Oduro, 1998
Fusarium oxysporum Schlecht.	BF BN CI GH GU ML NR NG SL SG TG	HI US VI GU PR	L, W	No	No	CABI, 2004
<i>Fusarium</i> <i>pallidoroseum</i> (Cooke) Sacc. = F. semitectum Berk. & Rav.	GH	US	F, L	No	Yes	Oduro, 1998
Irinopsis aciculosa	GH		L	Yes	No	Oduro, 1998

9 Leaf spot. If it is a scab, fruit can be culled through casual inspection.

Pest	West	US	Plant Part Affected ²	Quaranti ne	Follow Pathway	References
	Distribution ¹	Distribution	/ mooted	ne	-	
Lasiodiplodia	BF GM	US GU PR	F,Fw, L, S,	No	Yes	CABI, 2004
theobromae (Pat.)	GH GU		Sd			
Griffon & Maubl.	NG SG TG		A			
= Botryodiplodia						
theobromae Pat.,					*	
=						a
Diplodianatalensis						
Pole-Evans,	8					
teleomorph =						
Physalospora						· · · · ·
rhodina (Berkeley						
& Curtis) Cooke	CLON CUL			21	X.	CADL 2004
Leveillula taurica	CI GM GH	HI US PR	L, S	No	Yes	CABI, 2004
(Lev.) G. Arnaud	GUNK					
	NG SL SG					
	IG	LIC DD	L D C I C	NL	N	CADL 2004
Macrophomina	BF BN CI	USPR	L, K, Sd, S,	NO	NO	CABI, 2004
phaseolina (Tassi)	GMINK		W			
Gold	NG SL SG					
Nastria		LIC	LCDW	No	No	CADI 2004
harmatococca	ОП	05	L,5,K,W	INO	INO	CABI, 2004
Rark & Proomo						
Oidium	GH	US	LS	No	No	Farr at al 2004
ahelmoschi	UII	03	1,5	140	INU	1 all el ul., 2004
Thuem		3			⁶³ c	
Penicillium	NG	US	F	No	Ves	CABL 2004
digitatum (Pers	ing ing	05	1	110	103	0/101, 2004
Fr.) Sacc.					A	
Phomopsis	SG	US PR	F.Sd	No	Yes	CABL 2004
longicolla Hobbs		0.0111	1,04			01121, 2001
Pseudocercospora	GH	US	L	No	No	Farr et al., 1989:
abelmoschi (Ellis			~			MoFA, 2000
& Everh.)					A	
Deighton						
Puccinia sp.	NG		F	Yes	Yes	PIN. 2003
Pythium	CI GH ML	HI US PR	R, W	No	No	CABI, 2004
aphanidermatum	NG SL SG					,
(Edson) Fitzp.	TG					
Sclerotinia	NG	HIUS	F, Fw, L, R.	No	Yes	CABI, 2004
sclerotiorum			Sd. W			
(Lib.) de Bary						
Verticillium	NG	US	L, S, W	No	No	CABI, 2004
dahliae Kleb.						

Pest	West African Distribution ¹	US Distribution	Plant Part Affected ²	Quaranti ne	Follow Pathway	References
Virus				2		
Cotton leaf curl virus	BK BN CI GH NG NR TG		L	Yes	No	CABI, 2004; Oduro, 1998
Cucumber Mosaic Virus	CI GH NG SL TG	HI US PR	F, L, W	No	No	CABI, 2004
Hibiscus yellow vein mosaic	GH		L	Yes	No	Oduro, 1998
Okra leaf curl virus	GH		W	Yes	No	Brunt <i>et al.</i> , 1996+; N' Guessan <i>et al.</i> , 1992
Okra mosaic virus	NG			Yes		Vanlommel et al., 1996

Quarantine Pests Selected for Further Analysis.

Cryptophlebia leucotreta (Meyrick) (Lepidoptera: Tortricidae) Earias biplaga Walker (Lepidoptera: Noctuidae) Earias insulana (Boisduval) (Lepidoptera: Noctuidae) Helicoverpa armigera (Hubner) (Lepidoptera: Noctuidae) Spodoptera littoralis (Boisduval) (Lepidoptera: Noctuidae) Leucinodes orbonalis Guenee (Lepidoptera: Pyralidae)

5. Consequences of Introduction—Economic/Environmental Importance

Potential consequences of introduction are rated using five risk elements:

- 1. Climate-Host Interaction
- 2. Host Range
- 3. Dispersal Potential
- 4. Economic Impact
- 5. Environmental Impact

These elements reflect the biology, host ranges and climatic/geographic distributions of the pests. For each risk element, pests are assigned a rating of Low (1 point), Medium (2 points) or High (3 points) (USDA, 2000). A Cumulative Risk Rating is then calculated by summing all risk element values. The values determined for the Consequences of Introduction for each pest are summarized in Table 3.

The major sources of uncertainty in this risk assessment are similar to those in other risk assessments: the use of a developing process (Orr *et al.*, 1993; USDA, 2000), the approach used to combine risk elements (Bier, 1999; Morgan and Henrion, 1990), and the evaluation of risk by comparisons to lists of factors within the guidelines (Orr *et al.*, 1993). To address this last source of uncertainty, of factor lists were interpreted as illustrative and not exhaustive. Other traditionally recognized sources of uncertainty are the quality of the biological information (Gallegos and Bonano, 1993), which includes uncertainty whenever biological information is lacking on the regional flora and fauna. Inherent biological variation within a population of organisms also introduces uncertainty (Morgan and Henrion, 1990).

Risk Element #1- Climate-Host Interactions

If a species encounters suitable climate and hosts in the area where it is introduced, the organism may survive and achieve pest status in the new environment. This risk element is evaluated on the minimum number of U.S. "Plant Hardiness Zones" in which the species might achieve pest status (USDA, 1990). Risk ratings are based on the following criteria:

Low (1): the species is only likely to become established in one hardiness zone Medium (2): the species is likely to become established in two or three hardiness zones High (3): the species is likely to become established in four or more hardiness zones

Risk Element #2- Host Range

The risk posed by a plant pest depends on its ability to establish a viable, reproductive population and its potential to injure plants. For arthropods, risk is assumed to be positively correlated with host range. For pathogens, risk is assumed to depend on host range, aggressiveness, virulence and pathogenicity; for simplicity, risk is rated as a function of host range:

Low (1): pest attacks a single species or multiple species within a single genus Medium (2): pest attacks multiple species within a single plant family High (3): pest attacks multiple species among multiple plant families

Risk Element #3-Dispersal Potential

A pest may disperse after arriving in a new area. The following items are considered in regard to dispersal potential: reproductive patterns of the pest (*e.g.*, voltinism, biotic potential); inherent powers of movement; factors facilitating dispersal, wind, water, presence of vectors, humans, *etc*.

Low (1): pest has neither high reproductive potential nor rapid dispersal capability Medium (2): pest has either high reproductive potential OR the species is capable of rapid dispersal

High (3): Pest has high biotic potential, *e.g.*, many generations per year, many offspring per reproduction ("r-selected" species), AND evidence exists that the pest is capable of rapid dispersal, *e.g.*, over 10km/year under its own power; via natural forces, wind, water, vectors, *etc.*, or human-assistance.

Risk Element #4-Economic Impact

Introduced pests can cause a variety of direct and indirect economic impacts. These impacts are divided into three primary categories (other types of impacts may occur): lower yield of the host crop, *e.g.*, by causing plant mortality, or by acting as a disease vector; lower value of the commodity, *e.g.*, by increasing costs of production, lowering market price, or a combination; and loss of foreign or domestic markets due to the presence of a new quarantine pest.

Low (1): pest causes any one or none of the above impacts Medium (2): pest causes any two of the above impacts High (3): pest causes all three of the above impacts

Risk Element #5- Environmental Impact

A pest may cause significant, direct consequences to the environment, *e.g.*, cause an ecological disaster or reduce biodiversity. In the context of the National Environmental Policy Act (NEPA) (7CFR§372), significance is qualitative and encompasses the likelihood and severity of an environmental impact. The act describes an environmental pest as: "expected to have direct impacts on species listed by Federal Agencies as endangered or threatened (50CFR§17.11 and §17.12), by infesting/infecting a listed plant. If the pest attacks other species within the genus or other genera within the family, and preference/no preference tests have not been conducted with the listed plant and the pest, then the plant is assumed to be a host; pest is expected to have indirect impacts on species listed by Federal Agencies as endangered or threatened by disrupting sensitive, critical habitat; introduction of the pest would stimulate chemical or biological control programs."

Low (1): none of the above would occur Medium (2): one of the above would occur High (3): two or more of the above would occur.

Consequences of Introduction: Cryptophlebia leucotreta Meyrick			
(Lepidoptera: Tortricidae)			
Risk Element #1: Climate – Host Interaction			
Cryptophlebia leucotreta is distributed throughout Africa (CABI, 2004). Its	(2)		
occurrence corresponds with U.S. Plant Hardiness Zones 9-11 (USDA, 1990).			
Risk Element #2: Host Range	High		
There are more than 70 species identified as host species to C. leucotreta (CABI,	(3)		
2004). Primary species include Rutaceae (Citrus spp., Citrus sinensis),			
Malvaceae (Gossypium spp., Abelmoschus esculentus, Abutilon hybridum),			
Poaceae (Zea mays, Sorghum), Euphorbiaceae (Ricinus communis), Theaceae			
(Camellia sinensis), Lauraceae (Persea Americana), Myrtaceae (Psidium	1. ¹⁰		
guajava), Oxalidaceae (Averrhoa carambola), Bromeliaceae (Ananas comosus),			
Annonaceae (Annona muricata), Bombacaceae (Ceiba pentandra), Rubiaceae			
(Coffea Arabica), Solanaceae (Capsicum), Sapindaceae (Litchi chinensis),	й н		
Anacardiaceae (Mangifera indica), Oleaceae (Olea europaea subsp. europaea),			
Rosaceae (Prunus persica), Punicaceae (Punica granatum), and Proteaceae	· .		
(Macadamia spp., Macadamia ternifolia).			
Risk Element #3: Dispersal Potential	High		
Females can lay between 100 – 400 eggs overnight (CABI, 2004), and typically	(3)		
lay 12 eggs per fruit (Bedford et al., 1998). It is rare for them to lay more than			
20 eggs per fruit; however, 65 eggs have been observed on a single fruit			
(Bedford et al., 1998). Survival of the first instar is temperature dependent (low	v.		
winter temperatures can be lethal) (Bedford et al., 1998). The life cycle varies			
with the season, although C. leucotreta typically has 2-3 generations per year			
(Bedford et al., 1998). Adults are attracted to light (CABI, 2004). Larvae can be			
internationally transported via fruit, pods, inflorescence and cones (CABI, 2004).			

Risk Element #4: Economic Impact	High
<i>Cryptophlebia leucotreta</i> is a serious pest of South African citrus. Losses in late crop of cotton ranges between 42-90% in Uganda. (CABI, 2004). Host species include several important crops, and the damages caused by this species would be high once it is introduced and established in the United States. In 2002, U.S. cotton production was worth more than \$3593 million (NASS, 2003). In addition to cotton species, citrus and corn production for the year 2002 in U.S. Plant Hardiness Zones 9-12 valued at \$2605 million and \$1040 million, respectively (NASS, 2003). In South Africa, crop damages can be as high as 50% on citrus species (Bedford <i>et al.</i> , 1998).	(3)
Risk Element #5: Environmental Impact This species has a potential to attack Endangered and Threatened species, such as <i>Quercus hinckleyi</i> (TX) (USFWS, 2002). There are several controls available; however, it is difficult to establish effective controls because the moth	High (3)
has many alternative hosts (CABI, 2004). Introduction and establishment of <i>C. leucotreta</i> in the United States would stimulate chemical or biological control programs.	

Consequences of Introduction: Earias biplaga (Lepidoptera: Noctuidae)	Risk Value
Risk Element #1: Climate – Host Interaction	Medium ¹⁰
Earias biplaga occurs in USDA Plant Hardiness Zones 9-13, ranging throughout	(2)
Africa (CABI, 2004; USDA, 1990).	
Risk Element #2: Host Range	High
This insect has multiple hosts in the family Malvaceae, and others in the families	(3)
Bombacaceae and Sterculiaceae (CABI, 2004).	
Risk Element #3: Dispersal Potential	High
Noctuid moths are generally strong fliers capable of flying miles in one night	(3)
(Rochester et al., 2002). Females of E. biplaga lay from 100-400 eggs (CABI,	
2004).	1
Risk Element #4: Economic Impact	High
Earias biplaga is a pest of okra, cotton, and cocoa in Africa (CABI, 2004). This	(3)
insect has the potential to lower yields and increase the cost of production for	
U.S. cotton growers if it should become established in the United States. In	
2002, U.S. cotton production was worth more than \$3,593 million (NASS,	
1997).	
Risk Element #5: Environmental Impact	High
Earias biplaga feeds on hosts in the genus Hibiscus, which contain endangered species	(3)
in Hawaii. These endangered species are: Hibiscus arnottianus ssp. immaculatus, H.	
brackenridgei, H. clayi, and H. waimeae ssp. hannerae. Chemical controls are available	
to control E. biplaga. Introduction and establishment of E. biplaga may stimulate	
chemical controls in the United States.	

10 Although Plant Hardiness Zones cover more than 3 zones, climate-host interaction is rated medium because Plant Zones 9-13 are tropical zones.

Consequences of Introduction: Earias insulana (Lepidoptera: Noctuidae)	Risk Value		
Risk Element #1: Climate – Host Interaction	High		
Earias insulana occurs in USDA Plant Hardiness Zones 6-13, ranging	(3)		
throughout Africa, southern Europe, and southeast Asia (CABI, 2004).			
Risk Element #2: Host Range	High		
This insect feeds on multiple hosts in the families Malvaceae and Poaceae	(3)		
(CABI, 2004).			
Risk Element #3: Dispersal Potential	High		
Noctuid moths are generally strong fliers capable of flying miles in one night	(3)		
(Rochester et al., 2002). Females of E. insulana lay an average of 128.4 eggs in			
one study Anwar et al., 1973.	×.		
Risk Element #4: Economic Impact	High		
Earias insulana is a pest of cotton in Pakistan (Chamberlain et al., 1993), Egypt	(3)		
(Rashad and Ammar, 1984) and India (Dhawan et al., 1992) as well as being a			
pest of okra and other members of the family Malvaceae (CABI, 2004). This			
insect has the potential to lower yields and increase the cost of production for			
U.S. cotton growers if it should become established in the United States. In			
2002, U.S. cotton production was worth more than \$3,593 million (NASS,			
1997).			
Risk Element #5: Environmental Impact	High		
Earias insulana feeds on hosts in two genera, Abutilon and Hibiscus, which contain	(3)		
endangered species in Hawaii. These endangered species are: Abutilon eremitopetalum,			
A. menziesii, A. sandwicense, Hibiscus arnottianus ssp. immaculatus, H. brackenridgei,			
H. clayi, and H. waimeae ssp. hannerae. There are number of studies to examine			
methods and effectiveness of controls, such as cultural, nost-resistant, biological, and			
management programs were established. Introduction and establishment of $F_{introduction}$			
may stimulate chemical and biological controls in the United States			
, contract and crotogram controls in the control states.			

Consequences of Introduction: Helicoverpa armigera (Lepidoptera:	Risk Value
Noctuidae)	
Risk Element #1: Climate – Host Interaction	High
Climate-Host Interaction, This insect is widely distributed and known to occur in all	(3)
parts of Europe, Middle East, Central and South Asia, Far East, Africa, Australia, and	
Oceania (CABI, 2003). Establishment is possible in U.S. Plant Hardiness Zones 5-11.	v
Risk Element #2: Host Range	High
<i>Helicoverpa armigera</i> is polyphagous. It infests crop and non-crop hosts representing over 10 genera and over four families (Zhang, 1994). It is a major pest of cotton	(3)
(Gossypium spp.), pigeon pea (Cajanus cajan), chickpea (Cicer arietinum), tomato	
(Lycopersicum esculentum), sorghum (Sorghum spp.) and cowpea (Vigna unguiculata).	6 98
Other hosts include groundnut (Arachis hypogaea), okra (Abelmoschus esculentus), peas	
(Pisum sativum), soybeans (Glycine max), other legumes, tobacco (Nicotiana tabacum),	
potatoes (Solanum tuberosum), maize (Zea mays), flax (Linu.n usitatissimum), a number	
of fruits (Prunus spp. and Citrus spp.), forest trees and a range of vegetable crops	
(CABI, 2004).	

Risk Element #3: Dispersal Potential	High
As with other noctuids, the pest is capable of flying long distances of many miles to	(3)
disperse. Internal larvae may be dispersed long distances in fruits (CABI, 2004).	
Females may lay over 700 eggs during their lifetime and there may be up to six	
generations per year (CABI, 2004) and may produce two to six generations depending	
on the climatic conditions (Smith et al., 1997). Larvae have limited mobility, but adults	
are capable of flight (CABI, 2004; Smith et al., 1997).	
Risk Element #4: Economic Impact	High
Economic Impact, Larvae are major pests of tomato, maize, cotton, and other crops	(3)
(CABI, 2004), becoming major pests if they establish. For example, in India, losses of	
up to 50% of the potato crop have been recorded (CABI, 2004). As an A2 pest for	
Europe, establishment in the United States could lead to loss of export markets (EPPO,	
2003).	
Risk Element #5: Environmental Impact	Medium
A wide range of wild plant species support larval development of H. armigera. Among	(2)
others, larvae can feed on the genera Allium, Amaranthus, Helianthus, Helianthus,	
Prunus, Solanum, and Vigna (CABI, 2004), which contain threatened or endangered	
species (USFWS, 2002). Helicoverpa armigera is very similar to H. virescens in	
phylogeny and in behavior (Farrow and Daly, 1987). Heliothis virescens is widespread	
in the United States so it is unlikely that the introduction of the similar species, H.	
armigera, would have a large environmental impact.	

Consequences of Introduction: Spodoptera littoralis (Lepidoptera: Noctuidae)	Risk Value	
Risk Element #1: Climate – Host Interaction		
Climate-Host Interaction, This insect is found in Africa, southern Europe, and the	(3)	
Middle East (CABI, 2004). It could become established in U.S. Plant Hardiness Zones		
8-11.	5.	
Risk Element #2: Host Range	High	
The host range of S. littoralis covers over 40 families, containing at least 87 species of	(3)	
plants of economic importance (CABI, 2004). For example: cotton (Gossypium spp.),		
tobacco (Nicotiana tabacum), potato (Solanum tuberosum), tomato (Lycopersicum		
esculentum), onion (Allium cepa), citrus (Citrus spp.), beans (Phaseolus spp.), carrots		
(Daucus carota), peppers (Capsicum annuum), grapes (Vitis spp.), alfalfa (Medicago	10. 	
sativa) and various grasses (CABI, 2004).		
Risk Element #3: Dispersal Potential	High	
Noctuids can disperse over long distances (Farrow and Daly, 1987). Adult S. litttoralis	(3)	
fly at night, with a flight range of 1.5 km in a 4-hour period (CABI, 2004). In optimal		
climates, the pest can have up to 7 overlapping generations per year, with an average of		
20-1000 eggs produced by each female (CABI, 2004).		
Risk Element #4: Economic Impact	High	
Spodoptera littoralis is one of the most destructive agricultural lepidopterous pests	(3)	
within this subtropical and tropical range (CABI, 2004). It can attack numerous		
economically important crops throughout the year. It lowers crop yield, increases		
production costs, and will cause market loss as a new quarantine pest.		

Risk Element #5: Environmental Impact	Medium
Threatened and endangered and species of Allium, Solanum, Vigna, Amaranthus,	(2)
Prunus, Hibiscus, Trifolium and Quercus may be at risk since these genera are known to	
be hosts for S. littoralis. New control measures would be unlikely because the current	
practices in commercial agriculture address a complex of similar noctuid pests.	

Consequences of Introduction: Leucinodes orbonalis (Lepidoptera:	Risk Value	
Pyralidae)		
Risk Element #1: Climate – Host Interaction	Medium	
Climate-Host Interaction, This insect is found in sub-Saharan Africa and India	(2)	
corresponding to U.S. Plant Hardiness Zones 10-13 (CABI, 2004). Only zone 10 occurs		
in the southern extremes of the United States ({USDA, 1990 #5819}).		
Risk Element #2: Host Range	High	
Leucinodes orbonalis feeds primarily on hosts in the family Solanaceae, but has been	(3)	
recorded on plants in the families Convolvulaceae and Fabaceae (CABI, 2004)		
Risk Element #3: Dispersal Potential	High	
Suresh et al., ({, 1996 #12370}) recorded fecundity of 62 and 164 eggs per female, but	(3)	
Singh and Singh ({, 2001 #15335}) recorded an average of 174 eggs per female. Adult		
behavior has been little studied, but other moths in the family Pyralidae have been		
shown to be capable of medium to long range flight (Cherry and Wilson, 2005; Shirai,		
1998)		
Risk Element #4: Economic Impact	High	
This insect causes extensive damage to okra and eggplant, lowering the yield and	(3)	
marketability of the crops ({Frempong, 1979 #11860}{Youdeowei, 2002 #15482}).		
Risk Element #5: Environmental Impact	Medium	
Leucinodes orbonalis feeds on plants in the genus Solanum, which also contains the	(2)	
endangered species Solanum drymophilum in Puerto Rico and S. incompletum and S.		
sandwicense in Hawaii.		

For each pest, the sum of the five risk elements gives a Cumulative Risk Rating. This Cumulative Risk Rating is considered to be a biological indicator of the potential of the pest to establish, spread, and cause economic and environmental impacts. The summary of risk ratings for Consequences of Introduction is shown in Table 3.

Low: 5-8 points Medium: 9-12 points High: 13-15 points

Table 3. Risk Rating for Consequences of Introduction

Pest	Risk Element 1 Climate/Host Interaction	Risk Element 2 Host Range	Risk Element 3 Dispersal Potential	Risk Element 4 Economic Impact	Risk Element 5 Environmental Impact	Cumulative Risk Rating
Cryptophlebia leucotreta	High (3)	High (3)	High (3)	High (3)	High (3)	High (15)
Earias biplaga	Medium	High	High	High	High	High

20

	(2)	(3)	(3)	(3)	(3)	(14)
Earias insulana	High	High	High	High	High	High
	(3)	(3)	(3)	(3)	(3)	(15)
Helicoverpa armigera	High	High	High	High	Medium	High
	(3)	(3)	(3)	(3)	(2)	(14)
Spodoptera littoralis	High	High	High	High	Medium	High
	(3)	(3)	(3)	(3)	(2)	(14)
Leucinodes orbonalis	Medium	High	High	High	Medium	High
	(2)	(3)	(3)	(3)	(2)	(13)

6. Introduction Potential

Each pest is rated with respect to its Likelihood of Introduction, which is based on two separate components. First, an estimate is made concerning the quality of the commodity likely to be imported (Risk Element #6). Second, pest opportunity (Risk Element #7) is estimated using five biological features. Details of those two Risk Elements and their rating criteria are provided in USDA APHIS (2000); the ratings and cumulative score for Risk Element #6 and #7, i.e., the "Likelihood of Introduction Risk Rating" are shown in Table 4.

Risk Element #6: Pest Opportunity (Survival and Access to Suitable Habitat and Hosts)

For each pest, the following six sub-elements were considered:

1. Quantity of commodity imported annually:

The likelihood that an exotic pest will be introduced depends on the amount of potentially infested commodity that is imported. For qualitative pest risk assessments, the amount of commodity imported is estimated in units of standard 40 foot long shipping containers. In those cases where the quantity of a commodity imported is provided in terms of kilograms, pounds, number of items, etc., the number of units is converted the units into terms of 40 foot shipping containers.

Low (1 point): < 10 containers/year Medium (2 points): 10 – 100 containers/year High (3 points): > 100 containers/year

Total okra production in Ghàna in 2003 was 100,000 metric tons (FAO, 2005). Sea shipping containers which are 40 foot in length hold approximately 40,000 pounds (20 U.S. tons); this is used for various estimate of commodity shipment (USTRIS, 2005). Anticipated volume of okra to be exported from Ghana is unknown; however, high volume of okra (> 100 containers/year) is likely to be shipped into the United States. Therefore, Quantity of commodity imported annually is rated High (3).

2. Survive postharvest treatment:

For this sub-element, postharvest treatment refers to any manipulation, handling, or specific phytosanitary treatment to which the commodity is subjected. Examples of postharvest treatment include culling, washing, chemical treatment, cold storage, etc. If there is no postharvest treatment, the estimate the likelihood of this sub-element is High.

The false codling moth, *Cryptophlebia leucotreta*, the Egyptian stem borer, *Earias biplaga*, the spiny bollworm, *Earias insulana*, the cotton bollworm, *Helcoverpa armigera*, and *Spodoptera littoralis* are rated Medium. Okra will not be washed but inspected in the packing house to cull out malformed or damaged fruits. Therefore, all the insects are rated Medium.

3. Survive Shipment:

The shipping conditions of okra from Ghana are unknown, but would probably be short in duration owing to the perishable nature of fresh okra. All pests are rated High for this risk element.

4. Not be detected at the port of entry:

Unless specific protocols with special inspection of the commodity in question are in place, standard inspection protocols for like commodities are assumed. If no inspection is planned, estimate this sub-element as High.

All the species are rated Medium. Fruits are harvested early to avoid overly mature fruits and attack by the borers. The entry points in fruits are visible and could be detected by inspection.

5. Imported or move subsequently to an area with an environment suitable for survival: *Cryptophlebia leucotreta, Earias biplaga*, and *Leucinodes orbonalis* are rated Medium because they are tropical and subtropical species. Tropical and subtropical locations are limited in the United States; in the continental United States, those regions are limited to the South and the West Coast, which comprise an estimated 10-12% of the total land area of the continental United States.

Earias insulana, Helicoverpa armigera, and *Spodoptera littoralis* are rated High because suitable habitats contain not only subtropical and tropical zones but also temperate zones

6. Come into contact with host material suitable for reproduction:

Even if the final destination of infested commodities is conducive for pest survival, suitable host material must be available in order for the pest to survive. Consider the complete host range of the pest species.

Cryptophlebia leucotreta, Earias biplaga, E. insulana, Helicoverpa armigera, Leucinodes orbonalis and *Spodoptera littoralis* are rated High. All five pests have wide range of host species. *Cryptophlebia leucotreta* and *Helicoverpa armigera* are polyphagous species. *Earias species attack cotton, rice, and maize which are widely distributed throughout the United States.*

A summary of the ratings for Likelihood of Introduction is depicted in Table 4.

Low: 6-9 points Medium: 10-14 points High: 15-18 points

5	Subelement 1	Subelement 2	Subelement 3	Subelement 4	Subelement 5	Subelement 6	
Pešt	Quantity imported annually	Survive postharvest treatment	Survive shipment	Not detected at port of entry	Moved to suitable habitat	Contact with host material	Cumulative Risk Rating
Cryptophlebia	High	Medium	High	Medium	Medium	High	High
leucotreta	(3)	(2)	(3)	(2)	(2)	(3)	(15)
Earias biplaga	High	Medium	High	Medium	Medium	High	High
	(3)	(2)	(3)	(2)	(2)	(3)	(15)
Earias insulana	High	Medium	High	Medium	Medium	High	High
	(3)	(2)	(3)	(2)	(3)	(3)	(16)
Helicoverpa	High	Medium	High	Medium	High	High	High
armigera	(3)	(2)	(3)	(2)	(3)	(3)	(16)
Spodoptera	High	Medium	High	Medium	High	High	High
littoralis	(3)	(2)	(3)	(2)	(3)	(3)	(16)
Leucinodes	High	Medium	High	Medium	Medium	High	High
orbonalis	(3)	(2)	(3)	(2)	(2)	(3)	(15)

Table 4. Risk Rating for Likelihood of Introduction: (Risk Element #6)

C. Conclusion - Pest Risk Potential and Pests Requiring Phytosanitary Measures

To estimate the Pest Risk Potential for each pest, the Cumulative Risk Rating for the consequences of Introduction and the Cumulative Risk Rating for the Likelihood of Introduction are summed in Table 5. The Pest Potential rating is as follows:

Low: 11 – 18 points Medium: 19 – 26 points High: 27 – 33 points

Table 5. Summary of pest risk potential

Pest	Consequences of Introduction	Likelihood of Introduction	Pest Risk Potential	Risk Rate
Cryptophlebia leucotreta	High (15)	High (15)	30	High
Earias biplaga	High (14)	High (15)	29	High

Earias insulana	High (15)	High (16)	31	High
Helicoverpa armigera	High (14)	High (16)	30	High
Spodoptera littoralis	High (14)	High (16)	30	High
Leucinodes orbonalis	High (13)	High (15)	28	High

Pest Risk Potential ratings have the following suggested meanings (APHIS, 2000):

Low:	Pest will typically not require specific mitigation procedures. The port-of-
	entry inspection to which all imported commodities are subjected can be
	expected to provide sufficient phytosanitary security.
Medium:	Specific phytosanitary measures may be necessary.
High:	Specific phytosanitary measures are strongly recommended. Port-of-entry
	inspection is not considered sufficient to provide phytosanitary security.

As stated in the Guidelines (APHIS, 2000) detailed examination and choice of appropriate sanitary and phytosanitary measures to mitigate pest risk for commodities with particular pest risk potential scores or ratings is undertaken as part of the pest risk management phase and is not discussed in this document. The appropriate risk management strategy for a particular pest depends on the risk posed by that pest.

C. Authors

Bernard A. Boateng, Entomologist, University of Ghana Haruna Braimah, Entomologist, Crops Research Institute, Kumasi, Ghana **Mary Glover-Amengor, Biochemist, Food Research Institute, Accra, Ghana** Angela Osei-Sarfoh – Applied Biologist, Ghana Ministry of Food and Agriculture Ruth Woode, Entomologist, Ghana Ministry of Food and Agriculture Shawn Robertson – Entomologist, APHIS-IS Yu Takeuchi – Plant Physiologist, APHIS-PPQ

D. Literature Cited

- Anwar, M., Ashraf, M., and Arif, M. D. 1973. Mating, oviposition and gamma sterilization of the spotted bollworm of cotton, Earias insulana. Entomologia Experimentalis et Applicata 16: 478-482.
- APHIS. 2000. Guidelines for pathway-initiative pest risk assessments, v 5.02. Animal and Plant Health Inspection Service, United States Dept Agriculture. <<u>http://www.aphis.usda.gov/ppq/pra/commodity/cpraguide.pdf</u>> *last accessed* 10 January 2002.
- Bedford, E. C. G., van den Berg, M. A., and Villiers, E. A. d. 1998. Citrus Pests in the Republic of South Africa, second edition., 2nd ed. Agricultural Research Council, Institute for Tropical and Subtropical Crops, Nelspruit, South Africa.

- Ben-Dov, Y. 1993. A Systematic Catalogue of the Soft Scales Insects of the World. Sandhill Crane Press, Inc., Gainesville, Florida. pp.536.
- Ben-Dov, Y., Miller, D. R., and Gibson, G. A. P. 2004. ScaleNet.
 - <<u>http://www.sel.barc.usda.gov/scalenet/scalenet.htm</u>> *last accessed* 3 November 2004.
- Bier, V. M. 1999. Challenges to the acceptance of probabilistic risk analysis. Risk Analysis 19: 703-710.
- Bolland, H. R., Gutierrez, J., and Flechtmann, C. H. W. 1998. World catalogue of the spider mite family (Acari: Tetranychidae). Brill, Boston.
- Brunt, A. A., Crabtree, K., Dallwitz, M. J., Gibbs, A. J., Watson, L., and Zurcher, E. J. 1996+. Plant Viruses Online: Descriptions and Lists from the VIDE Database. Version: 20th August 1996.' Australian National University, Research School of Biological Sciences. <<u>http://image.fs.uidaho.edu/vide/refs.htm</u>> *last accessed* February 2004.
- CABI. 2004. Crop Protection Compendium (2004 edition). CAB International, Wallingford, UK.,
- Chamberlain, D. J., Ahmad, Z., Attique, M. R., and Chaudhry, M. A. 1993. The influence of slow release PVC resin pheromone formulations on the mating behaviour and control of the cotton bollworm complex (Lepidoptera: Gelechiidae and Noctuidae) in Pakistan. Bulletin of Entomological Research 83: 335-343.
- Cherry, R., and Wilson, A. 2005. Flight activity of tropical sodworms (Lepidoptera: Pyralidae). Florida Entomologist 88.
- Cobbinah, J. R., and Osei-Owusu, K. 1988. Effects of neem seed extracts on insect pests of eggplant, okra and cowpea. Insect Science and its Application 9: 601-607.
- Dhawan, A. K., Srimannarayana, G. S., Simwat, G. S., and Nagaiah, K. 1992. Management of cotton pests on upland cotton Gossypium hirsutum with Navneem 95EC. Proceedings First National Symposium Allelopathy in agroecosystems agriculture and forestry, February 12 14, 1992 held at CCS Haryana Agricultural University, Hisar 125 004, India. 1992; 156 157.
- EPPO. 2003. EPPO Standard PM1/2(12) A1 and A2 Quarantine Lists. European Plant Protection Organization. Paris, France.

<<u>http://www.eppo.org/QUARANTINE/lists.html</u>> *last accessed* 3 December 2003.

- FAO. 1996. International Standards for Phytosanitary Measures, Guidelines for Pest Risk Analysis. Secretariat of the International Plant Protection Convention of the Food and Agriculture Organization (FAO) of the United Nations, Rome, pp. 18. I:\PERAL\DocumentLibrary\Shawn'sReferences\FAO 1996 ISPM 11.pdf
- FAO. 2005. FAOSTAT. http://faostat.fao.org/ last accessed June 2005.
- Farr, D. F., Bills, G. F., Chamuris, G. P., and Rossman, A. Y. 1989. Fungi on Plants and Plant Products in the United States. APS Press, St. Pau., MN. pp.152.
- Farr, D. F., Rossman, A. Y., Palm, M. E., and McCray, E. B. 2004. Fungal Databases. Systematic Botany & Mycology Laboratory, ARS, USDA. <<u>http://nt.ars-grin.gov/fungaldatabases/>last accessed</u> December 2003.
- Farrow, R. A., and Daly, J. C. 1987. Long-range movements as an adaptive strategy in the genus *Heliothis* (Lepidoptera: Noctuidae): a review of its occurrence and detection in four pest species. Australian Journal of Zoology 35: 1-24.
- Gallegos, D. P., and Bonano, E. J. 1993. Consideration of uncertainty in the performance assessment of radioactive waste disposal from an international regulatory perspective. Relaiability Engineering and System Safety 42: 111-123.

- Gonzalez-Chavira, M., Rodriguez-Guerra, R., Hernandez-Godinez, F., Acosta-Gallegos, J. A., Martinez-de-la-Vega, O., and Simpson, J. 2004. Analysis of pathotypes of *Colletotrichum lindemuthianum* found in the central region of Mexico and resistance in elite germ plasm of *Phaseolus vulgaris*. Plant Disease 88: 152-156.
- Gunn, C. R., and Ritchie, C. 1982. Report of the Technical Committee to Evaluate Noxious Weeds; Exotic Weeds for Federal Noxious Weed Act (unpublished).
- Hill, D. S. 1994. Agricultural Entomology. Timber Press, Portland, Oregon. pp.640.
- Holm, L. G. 1997. World weeds: natural histories and distribution. John Wiley & Sons, N.Y. pp.1129.
- Holm, L. G., Pancho, J. V., Herberger, J. P., and Plucknett, D. L. 1979. Geographical atlas of world weeds. John Wiley and Sons, N.Y. pp.391.
- Hopper, B. E. 1995. Compendium of Phytosanitary Terms. North American Plant Protection Organization, Nepean, Ontario, Canada,
- Leston, D. 1972. The natural history of some West African insects. Entomologist's Monthly Magazine 108: 110-122.
- MoFA. 2000. Handbook of crop protection recommendations in Ghana: an IPM approach, Vol. 2: vegetables. Ghana Ministry of Food and Agriculture-Plant Protection and Regulatory Services Directorate, Pokuase, Ghana, pp. 92.
- Morgan, M. G., and Henrion, M. 1990. Uncertainty. Cambridge Univ. Press, United Kingdom. pp.332.
- N' Guessan, K., Fargette, D., Fauquet, C., and Thouvenel, J. C. 1992. Aspects of the epidemiology of okra leaf curl virus in Cote d'Ivoire. Tropical Pest Management 38: 122-126.
- NASS. 1997. Census of agriculture: ranking of states and counties. National Agricultural Statistics Service, United States Department of Agriculture, vol. 2, subject series part 2. <<u>http://www.nass.usda.gov/census/census97/rankings/ac97s-3r.pdf</u>> *last accessed* March 2004.
- NASS. 2003. Census of Agriculture. National Agricultural Statistics Service, United States Department of Agriculture. <<u>http://www.nass.usda.gov/census/</u>> *last accessed* December 2003.
- Obeng-Ofori, D., and Sackey, J. 2003. Field evaluation of non-synthetic insecticides for the management of insect pests of okra Abelmoschus esculentus (L.) Moench in Ghana. Sinet, Ethiopian Journal of Science 26: 145-150.
- Oduro, K. A. 1998. Checklist of plant diseases in Ghana. Ghana Ministry of Food and Agriculture-Plant Protection and Regulatory Services Directorate, Accra, Ghana. pp.105.
- Orr, R. L., Cohen, S. D., and Griffin, R. L. 1993. Generic non-indigenous pest risk assessment process: "The generic process" (For estimating pest risk associated with the introduction of non-indigenous organisms). USDA-APHIS, Policy and Program Development, MD, pp. 40.
- PIN. 2003. Port Interception Network. Pest interception records. Animal and Plant Health Inspection Service of the United States Department of Agriculture,
- Pogue, M. 2004. Moths of North America, No Common Name (*Anomis flava*). United States Geographical Survey, Northern Prairie Wildlife Research Center. <<u>http://www.npwrc.usgs.gov/resource/distr/lepid/moths/usa/9.htm</u>> *last accessed* August, 2004.

Rashad, A. M., and Ammar, E. D. 1984. Mass rearing of the spiny bollworm Earias insulana (Boisd.) on semi artificial diet. Bulletin de la Societe Entomologique d'Egypte: 239-244.

Reed, C. F. 1977. Economically important foreign weeds, potential problems in the United States, pp. 746, *In* Agriculture Handbook No. 498. Agricultural Research Service, Animal and Plant Health Inspection, U.S. Dept. of Agriculture, Washington, D.C.

Rochester, W. A., Zalucki, M. P., Ward, A., Miles, M., and Murray, D. A. H. 2002. Testing insect movement theory: empirical analysis of pest data routinely collected from agricultural crops. Computers and Electronics in Agriculture 35: 139-149.

Shirai, Y. 1998. Laboratory evaluation of flight ability of the Oriental corn borer Ostrinia furnacalis (Lepidoptera: Pyralidae). Bulletin of Entomological Research 88: 327-333.

- Smith, I. M., McNamara, D. G., Scott, P. R., and Holderness, M. 1997. Quarantine pests for Europe, 2nd Edition. CAB International., Wallingford, UK. pp.1425.
- USDA. 1990. United States National Arboretum, web version of USDA Plant Hardiness Zone Map. United States Department of Agriculture-Agricultural Research Service (ARS). Miscellaneous Publication No. 1475. USDA-ARS, Washington, D.C. <<u>http://www.usna.usda.gov/Hardzone/ushzmap.html></u> *last accessed*

USDA. 2000. Guidelines for Pathway-Initiated Pest Risk Assessments, Version 5.02. USDA-Animal and Plant Health Inspection Service. http://www.aphis.usda.gov/ppg/pra/commodity

USDA APHIS. 2000. Guidelines for Pathway-Initiated Pest Risk Assessments, Version 5.02. United States Department of Agriculture, Animal and Plant Health Inspection Service. <<u>http://www.aphis.usda.gov/ppq/pra/commodity/cpraguide.pdf</u>> *last accessed* 15 November 2003.

USFWS. 2002. Threatened and endangered species system (TESS). U. S. Fish and Wildlife Service. <<u>http://endangered.fws.gov/> last accessed</u> February 2004.

- USTRIS. 2005. United States Trade Internet System. <<u>http://www.fas.usda.gov/ustrade/> last</u> accessed 6 June 2005.
- Vanlommel, S., Duchateau, L., and Coosemans, J. 1996. The effect of okra mosaic virus and beetle damage on yield of four okra cultivars. African Crop Science Journal 4: 71-77.
- WSSA. 1989. Composite List of Weeds. Weed Science Society of America. pp.112.
- Zhang, B. C. 1994. Index of Economically Important Lepidoptera. CAB International, Wallingford, U.K. pp.599.